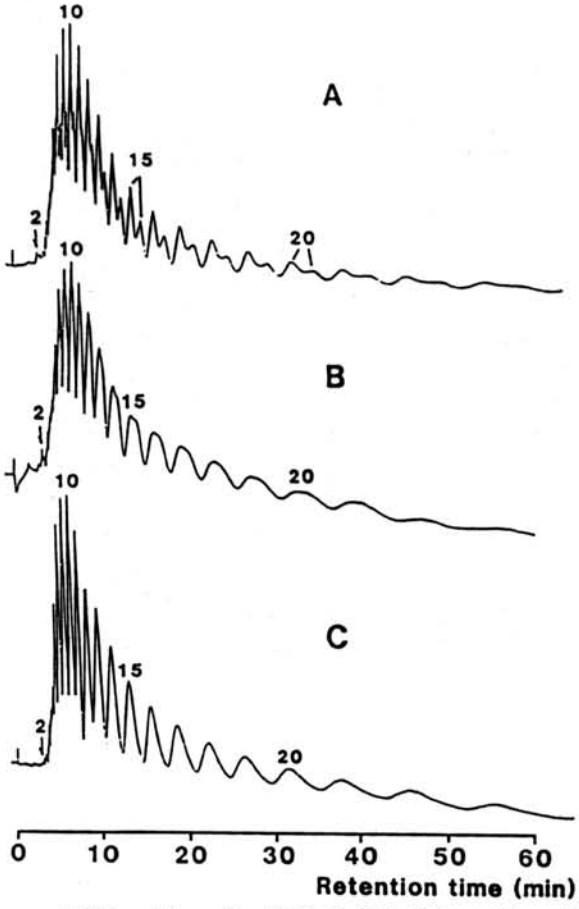
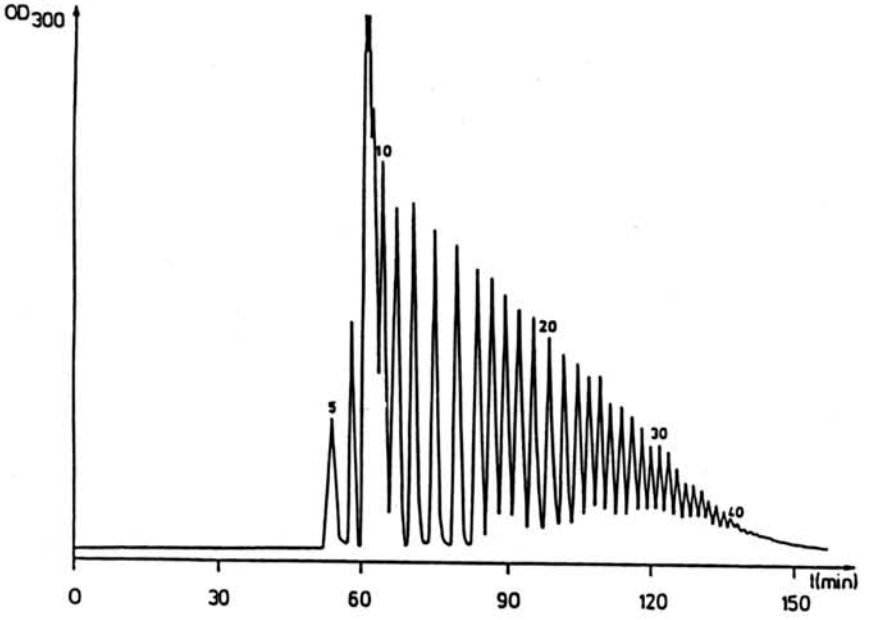
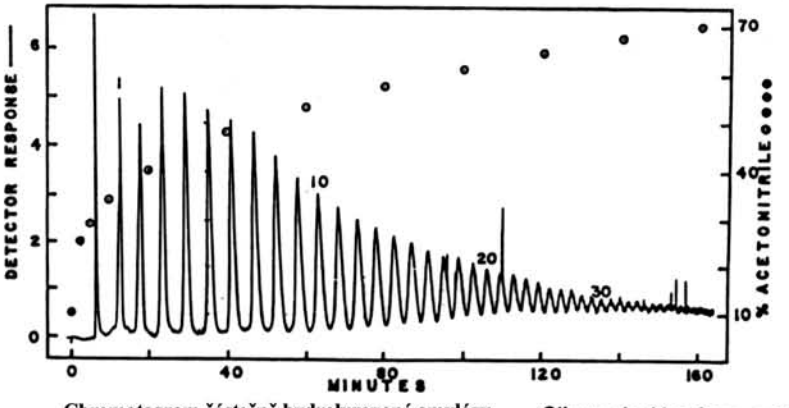


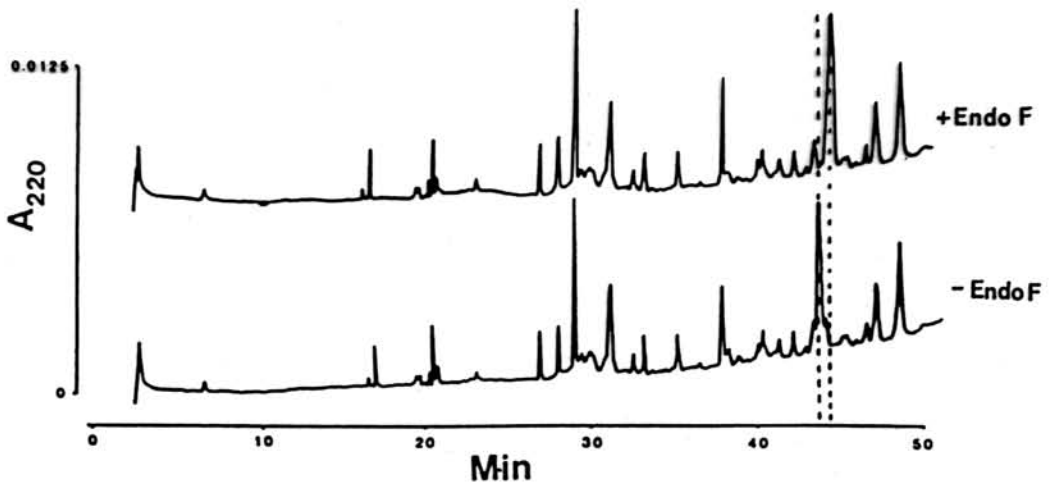
Eluční profily amylózy EX-1 s krátkým řetězcem (dp ≈ 17) na a Asahipak ODP-50 column (150 × 60 mm i.d.) (A) čistou vodou , (B)roztokem hydroxidu sodného pH 10, and (C) sodium hydroxide solution, pH 11. pH Measured at the column exit: (A) 6.28; (B) 9.50; (C) 10.75. The number on each peak indicates its dp. Chromatographic conditions: flow-rate, 1 ml/min; detector, Shodex RI SE-31 at 1×10^{-5} refractive index units full scale; temperature, ambient.



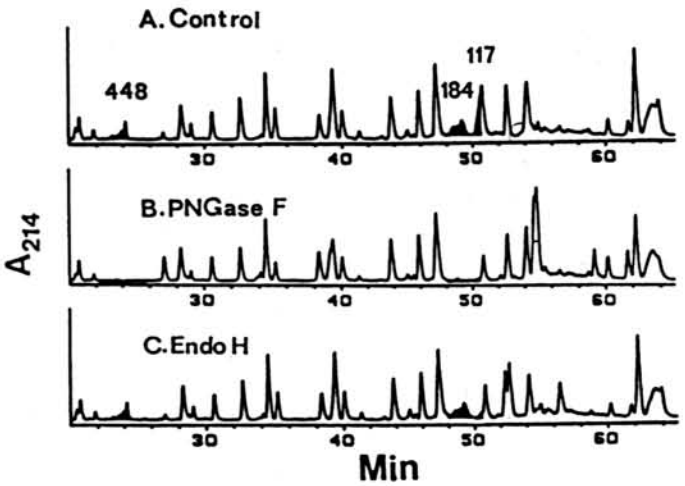
HPLC eluční profil 4-nitrophenyl-α-D-maltooligomerů na ODS Hypersil column, 3 μm, 4 × 250 mm. Mobile phase, water–methanol, linear gradient 96:90, 180 min.



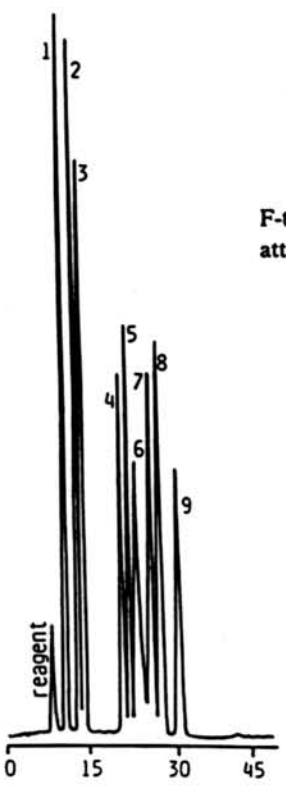
Chromatogram částečně hydrolyzované amylózy. Oligosaccharides from a 15-min acid hydrolyzate of amylose were acetylated and then chromatographed in a gradient of acetonitrile from 10 to 70% (v/v) in water over 160 min at a flow-rate of 1 ml/min. The degree of polymerization is indicated by numbers over the peaks.



Porovnání HPLC peptidových map nemodifikovaného a endo F-modifikovaného glykoproteinu. In this figure, 25 pmol of the peptide mixture resulting from the tryptic digestion of untreated and endo F-treated glycoprotein were applied in the digestion buffer to a Vydac C_{18} column (250 × 4.6 mm i.d.) equilibrated in 20 mM sodium phosphate, pH 2.5. The chromatograms were developed with a gradient of 0–45% acetonitrile over 45 min, followed directly with a 5 min gradient of 45–70% acetonitrile. The flow-rate was 1 ml/min and the column effluent was monitored at 220 nm. As seen in this figure, there is one peak in the chromatogram (highlighted by a dashed line at ca. 45 min of the profile) which exhibits an increase in its retention time following endo F treatment of the glycoprotein.

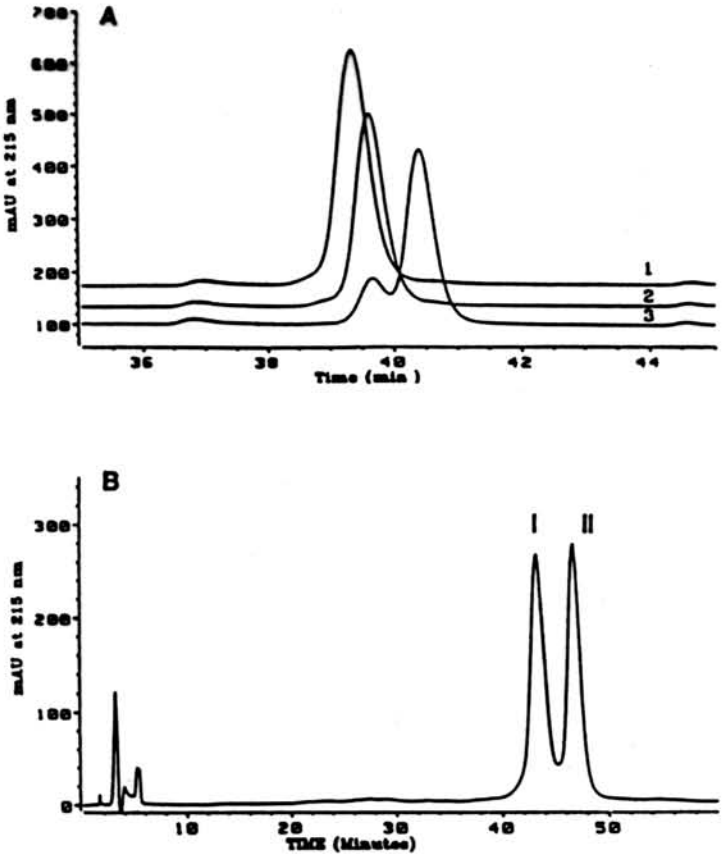


Porovnání tryptických map glykoproteinu rt-PA po štěpení glykosidázami. (A) control; (B) PNGase F-treated; (C) endo H-treated. Glycopeptide peaks are shaded and labeled by residue number of the attachment Asn.

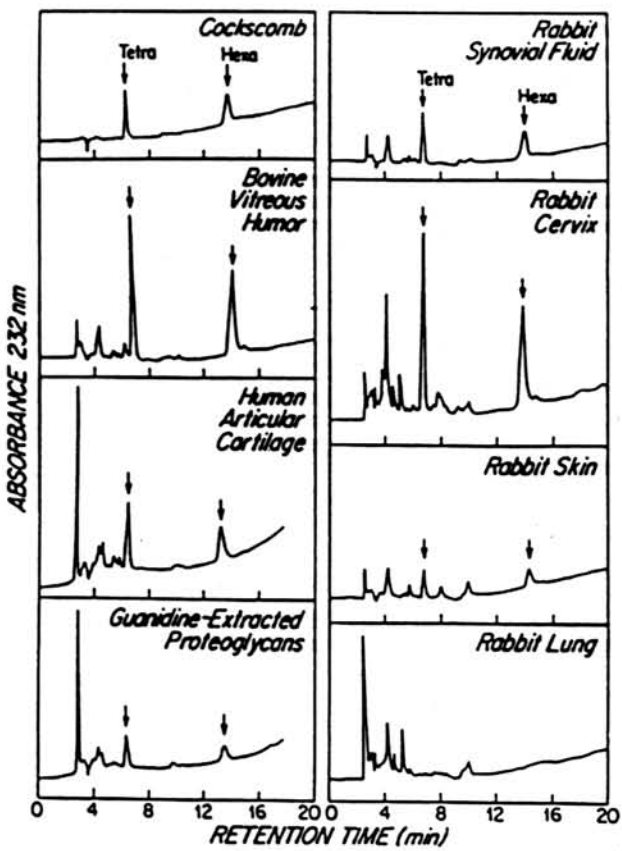


Dělení PMP (1-fenyl-3-methyl-5-pyrazolon) derivátů monosacharidů pomocí RP-HPLC.

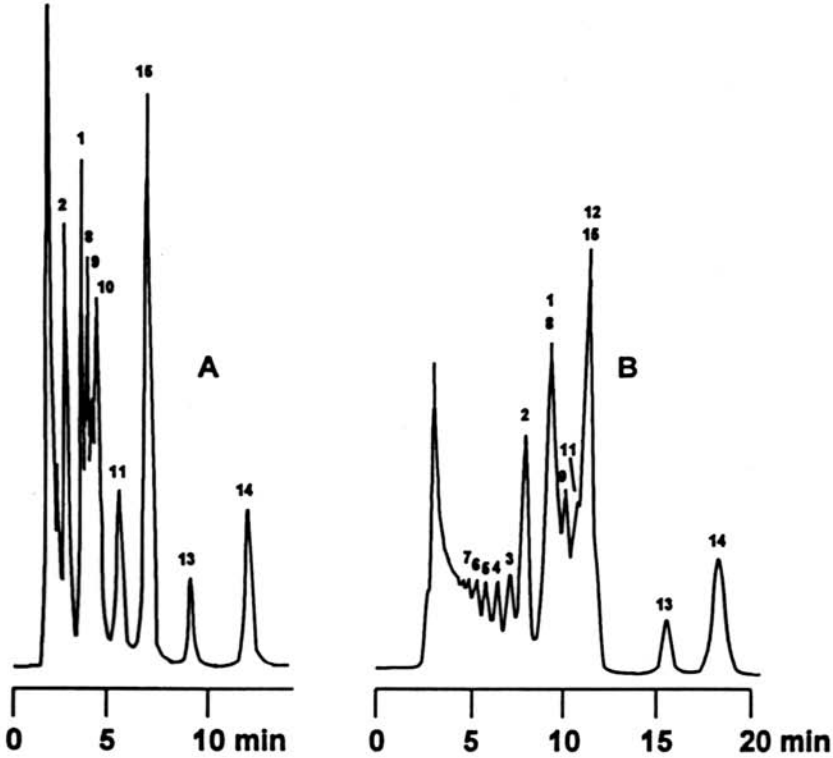
Eluent, 18% (v/v) acetonitrile in 0.1 M phosphate buffer, pH 7.0. Column, Capcell Pak C_{18}, 250 mm length, 4.6 mm i.d.; flow-rate, 1.0 ml/min; wavelength for detection 245 nm. Peak assignment: *1* = mannose; *2* = lyxose; *3* = rhamnose; *4* = *N*-acetylglucosamine; *5* = glucose; *6* = *N*-acetylgalactosamine; *7* = galactose; *8* = arabinose; *9* = fucose.

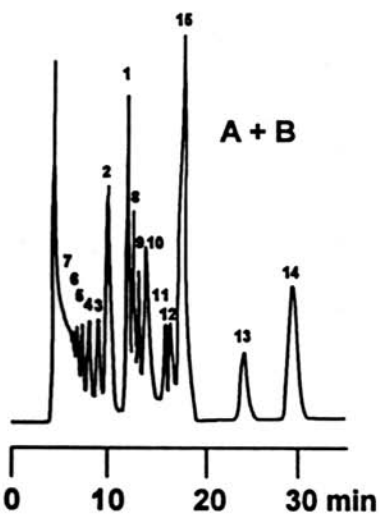


(A) HPLC na obrácené fázi G-CSFů. Chromatograms 1–3 (25 μg each): CHO-rhG-CSF; asialo CHO-rhG-CSF treated with neuraminidase; and CHO-rhG-CSF treated with neuraminidase and *O*-glycanase, respectively. (B) Cation-výměnná HPLC CHO-rhG-CSF (50 μg injected).



Iontově-párující RP-HPLC eluční profily hyaluronátových oligosacharidů připravených z různých tkání. All the chromatograms were run at the same sensitivity setting (0.5 absorbance units/full linear scale on the reporting integrator) and so are directly comparable.
Iontový pár: tetrabutylammonium fosfát





1...D-glucose
2...DP 2
3...DP 3
4...DP 4
5...DP 5
6...DP 6
7...DP 7
8...D-xylose
9...D-fructose
10...D-arabinose
11...dihydroxyacetone
12...1,5-anhydro-ß-D-glucose
13...hydroxymethylfurfural
14...furfural
15...ethylalcohol

Dělení mono- a oligosacharidů pomocí spojeních (coupled) kolon. Kolony: (A) Ca2+ kation-výměňná Spherogel, Carbohydrate N, Beckman; (B) Ag+ kation-výměnná, HPX 42A, Bio-Rad.